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(54) Title: COMPOSITIONS AND METHODS FOR IMMOBILIZING NUCLEIC ACIDS TO SOLID SUPPORTS

(57) Abstract

Compositions containing at least one bead conjugated to a solid support and further conjugated to at least one macromolecule, such as a nucleic acid and methods for making the compositions are provided. The resulting surfaces formed from the beads linked to the solid support advantageously provide increased surface area for immobilization of biological particles or macromolecules, particularly nucleic acids compared to "flat" surfaces. Furthermore, by selecting a bead with the desired functionality, an appropriate functionalization can be produced.

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COMPOSITIONS AND METHODS FOR IMMOBILIZING NUCLEIC ACIDS TO SOLID SUPPORTS

RELATED APPLICATIONS

For U.S. purposes, this application is a continuation-in-part of U.S.

5 application Serial No. 08/933,792, filed September 19, 1997, entitled
"COMPOSITIONS AND METHODS FOR IMMOBILIZING NUCLEIC ACIDS
TO SOLID SUPPORTS", which is a continuation-in-part of U.S.
application Serial No. 08/746,036 filed November 6, 1996, entitled
"BEAD LINKERS FOR IMMOBILIZING NUCLEI ACID TO SOLID
10 SUPPORTS". This application is also a continuation-in-part U.S.
application Serial No. 08/746,036.

For international purposes benefit of priority to each of these
applications is claimed herein. Benefit of priority is also claimed to U.S.
application Serial Nos. 08/786,988 and 08/787,639, each filed on
15 January 23, 1997.

Where permitted, the subject matter of each of U.S. application
Serial Nos. 08/933,792, 08/746,036, 08/786,988 and 08/787,639 are
herein incorporated in their entirety by reference thereto.

BACKGROUND OF THE INVENTION

20 In the fields of molecular biology and biochemistry, as well as in
the diagnosis of diseases, nucleic acid hybridization has become a
powerful tool for the detection, isolation, and analysis of specific
oligonucleotide sequences. Typically, such hybridization assays utilize an
oligodeoxynucleotide probe that has been immobilized on a solid support;
25 as for example in the reverse dot blot procedure (Saiki *et al.* (1989) *Proc*

1988) *probes attached to a solid surface have been developed for sequencing by hybridization (SBH) (Drmanac *et al.* (1989) *Genomics* 4:114-128), (Strezoska *et al.* (1991) *Proc. Natl. Acad. Sci.**

Acids Res. 22:2121-2125; Eggers et al. (1994) BioTechniques 17:516-524) or amino groups (see, e.g., Rasmussen et al. (1991) Anal. Biochem. 198:138-142). Although many of these methods were successful for their respective applications, when used to link nucleic acids to two-dimensional (flat) supports, the density of the immobilized oligodeoxynucleotide is often insufficient for the ensuing analyses (see, e.g., Lamture (1994) Nucl. Acids Res. 22:2121-2125; Eggers et al. (1994) BioTechniques 17:516-524).

Thus, there is a need for improved methods for immobilization that 10 provide higher densities of linked molecules for ensuing analyses. Therefore, it is an object herein to provide methods for preparing solid supports containing high densities of immobilized molecules, particularly nucleic acid molecules.

SUMMARY OF THE INVENTION

15 Compositions containing at least one bead conjugated to a solid support and further conjugated to at least one molecule, particularly a nucleic acid are provided. The bead is formed from any suitable matrix material known to those of skill in the art, including those that are swellable and nonswellable. The solid support is any support known to 20 those of skill in the art for use as a support matrix in chemical syntheses and analyses.

Preferably the bead is made of a material selected from materials that serve as solid supports for synthesis and for assays including but not limited to: silica gel, glass, magnet, polystyrene/1% divinylbenzene 25 resins, such as Wang resins, which are Fmoc-amino acid-4-(hydroxyl

chlorotriyl (2-chlorotriylchloride copolystyrene DVB resin) resin, Merrifield (chloromethylated copolystyrene-DVB) resin metal, plastic, cellulose, cross-linked dextrans, such as those sold under the tradename

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The above and further features and advantages of the compositions and methods provided herein will be elucidated in the following Figures, Detailed Description and Claims.

BRIEF DESCRIPTION OF THE DRAWINGS

5 Figure 1 is a schematic showing the covalent attachment of a bead to a solid support and DNA to the bead.

Figure 2 is a schematic showing the covalent attachment of Wang resin beads (p-benzyloxybenzyl alcohol copolystyrene-divinyl benzene (DVB) resin) to a solid support as described in Example 1.

10 Figure 3 is a schematic representation of nucleic acid immobilization via covalent bifunctional trityl linkers as described in Example 2.

Figure 4 is a schematic representation of nucleic acid immobilization via hydrophobic trityl linkers as described in Example 3.

15 Figure 5 shows a MALDI-TOF mass spectrum of a supernatant of the matrix treated Dynabeads containing bound oligo (5' iminobiotin-TGCACCTGACTC, SEQ. ID. No. 1). An internal standard (CTGTGGTCGTGC, SEQ. ID. No. 2) was included in the matrix.

20 Figure 6 shows a MALDI-TOF (matrix-assisted laser desorption/ionization (MALDI)-time-of-flight (TOF)) mass spectrum of a supernatant of biotin treated Dynabeads containing bound oligo (5' iminobiotin-TGCACCTGACTC, SEQ. ID. No. 1). An internal standard (CTGTGGTCGTGC, SEQ. ID. No. 2) was included in the matrix.

25 Figure 7 schematically depicts conjugation of an unextended primer to a bead via reaction of a 2', 3'-diol on the primer with horseradish peroxidase.

Figure 8 schematically depicts a pin tool apparatus.

Figure 9 depicts various pin conformations. Figures 9A shows a solid pin with a straight head. Figure 9B shows a solid pin with a

DETAILED DESCRIPTION OF THE INVENTION**A. Definitions**

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly understood by one of skill 5 in the art to which this invention belongs. All patents and publications referred to herein are, unless noted otherwise, incorporated by reference in their entirety. In the event a definition in this section is not consistent with definitions elsewhere, the definition set forth in this section will control.

10 As used herein, a molecule refers to any molecule or compound that is linked to the bead. Typically such molecules are macromolecules or components or precursors thereof, such as peptides, proteins, small organics, oligonucleotides or monomeric units of the peptides, organics, nucleic acids and other macromolecules. A monomeric unit refers to one 15 of the constituents from which the resulting compound is built. Thus, monomeric units include, nucleotides, amino acids, and pharmacophores from which small organic molecules are synthesized.

20 As used herein, macromolecule refers to any molecule having a molecular weight from the hundreds up to the millions. Macromolecules include peptides, proteins, nucleotides, nucleic acids, and other such 25 molecules that are generally synthesized by biological organisms, but can be prepared synthetically or using recombinant molecular biology methods.

As used herein, a biological particle refers to a virus, such as a 25 viral vector or viral capsid with or without packaged nucleic acid, phage

nucleotide acid, a single cell, including eukaryotic and prokaryotic cells or fragments thereof, a liposome or micellar agent or other packaging particle, and other such biological materials. For purposes herein,

In preferred embodiments, the beads are functionalized for the immobilization of nucleic acids and are stably associated with a solid support. Figure 1 depicts a bead conjugated to a solid support through one or more covalent or non-covalent bonds. Nucleic acids can be

5 immobilized on the functionalized bead before, during or after the bead is conjugated to the solid support.

Preferred nucleic acids for use herein are derivatized to contain at least one reactive moiety. Preferably the reactive moiety is at the 3' or 5' end. Alternatively, a nucleic acid can be synthesized with a modified 10 base. In addition, modification of the sugar moiety of a nucleotide at positions other than the 3' and 5' position through conventional methods is contemplated. Also, nucleic acid bases can be modified, e.g., by using N7- or N9- deazapurine nucleosides or by modification of C-5 of dT with a linker arm (see, e.g., Eckstein, ed., "Oligonucleotides and Analogues: A 15 Practical Approach," IRL Press (1991)). Alternatively, backbone-modified nucleic acids (e.g., phosphoroamidate DNA) can be used so that a reactive group can be attached to the nitrogen center provided by the modified phosphate backbone.

In preferred embodiments, modification of a nucleic acid, e.g., as 20 described above, does not substantially impair the ability of the nucleic acid or nucleic acid sequence to hybridize to its complement. Thus, any modification should preferably avoid substantially modifying the functionalities of the nucleic acid which are responsible for Watson-Crick base pairing. The nucleic acid can be modified such that a non-terminal 25 reactive group is present, and the nucleic acid when immobilized to the

hairpin structure having a duplex region.

Conjugation

The biological particles and macromolecules, such as nucleic acid molecules can be attached directly to the beads via a linker. Conjugation can be through any suitable means, particularly covalent or non-covalent attachment. For example, in one embodiment for conjugating nucleic acids to beads, the conjugating means introduces a variable spacer between the beads and the nucleic acids. In another embodiment, the conjugation is directly cleavable, such as a photocleavable linkage (e.g. streptavidin- or avidin- to biotin interaction can be cleaved by a laser, for example for mass spectrometry) or indirectly via photocleavable linker (see, e.g., U.S. Patent No. 5,643,722) or acid labile linker, heat sensitive linker, enzymatically cleavable linker or other such linker.

Similarly the bead is conjugated to the solid support by any suitable means, including those discussed herein for attachment of nucleic acids to beads. Thus, any of the conjugation methods and means discussed below with reference to conjugation of nucleic acids to beads can be applied for conjugation of beads to the solid support. In addition, it is understood that nucleic acids are exemplary of the molecules that can be conjugated to beads.

20 Conjugation via Linkers

Appropriate cross-linking agents for use for conjugating molecules to supports and beads and/or the beads to the supports include a variety of agents that are capable of reacting with a functional group present on a surface of the bead, insoluble support and or molecule, such as nucleic acid, and with a functional group present in the molecule, such as the

reactivity include homo- and hetero-bifunctional reagents, many of which are known in the art. Heterobifunctional reagents are preferred. A preferred bifunctional cross-linking agent is N-succinimidyl(4-iodoacetyl)

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e.g., formic acid, trifluoracetic acid, etc.) to ensure that the nucleic acid is cleaved and can be removed. In which case, the nucleic acid may be deposited as a beadless patch at the bottom of a well in the solid support or on the flat surface of the solid support. After addition of matrix

5 solution, the nucleic acid can then be desorbed, for example into a mass spectrometer.

Hydrophobic trityl linkers can also be exploited as acid-labile linkers by using a volatile acid or an appropriate matrix solution (e.g. a matrix solution containing, for example, 3-hydroxypicolinic acid (3-HPA)

10 to cleave the aminolink trityl group from the nucleic acid molecule). Also, the acid lability can be changed. For example, trityl, monomethoxy, dimethoxy- or trimethoxytrityl can be changed to the appropriate p-substituted and even more acid labile tritylamine derivatives of the nucleic acids (i.e. trityl ether and tritylamine bonds to the nucleic

15 acid can be made). Therefore, the nucleic acid may be removed from the hydrophobic linker, for example, by disrupting the hydrophobic attraction or by cleaving tritylether or tritylamine bonds under acidic or the usual mass spectrometry conditions (e.g. wherein the matrix, such as 3-HPA acts as an acid).

20 In a particularly preferred embodiment the bead is conjugated to the solid support and/or the nucleic acid is conjugated to the bead using an acid-labile bond. For example, use of a trityl linker, as further described in the following Examples 2 and 3, can provide a covalent or hydrophobic conjugation. Regardless of the nature of the conjugation,

25 the trityl group is readily cleaved in acidic conditions

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complementary nucleic acid, antibody-antigen and other such pairs known to those of skill in the art.

Examples of preferred binding pairs or linker/interactions is shown in the following Table 1:

5

TABLE 1

10

| LINKER/INTERACTION | EXAMPLES |
|---|---|
| streptavidin-biotin ^{a,c} /photolabile biotin ^b | biotinylated pin, avidin beads, photolabile biotin DNA |
| hydrophobic ^a | C18-coated pin, tritylated DNA |
| magnetic ^a | electromagnetic pin, streptavidin magnetic beads (e.g., DYNABEADS), biotin DNA |
| acid-labile linker ^b | glass pin, bifunctional trityl-linked DNA |
| amide bond(s) ^c | silicon wafer, Wang resin, amino-linked DNA |
| disulfide bond ^a | silicon wafer, beads are bound on the flat surface forming arrays or in arrays of nanoliter wells, thiol beads, thiolated DNA |
| photocleavable bond/linker | biotinylated pin/wafer, avidin beads, photolabile biotin DNA |
| thioether bond ^c | silicon wafer, beads are bound on the flat surface forming arrays or in arrays of nanoliter wells, thiolated DNA |

^aThese interactions are reversible. ^bThese non-reversible interactions are rapidly cleaved. ^cUnless cleavable-linkers are incorporated at some point in the scheme, only the complement of the solid-bound DNA can be analyzed in these schemes. ^dBeads are bound on the

15

In a particularly preferred embodiment the bead is conjugated to the solid support and/or the nucleic acid is conjugated to the bead using

methods in the following Examples 2 and 3, can provide a covalent or hydrophobic conjugation. Regardless of the nature of the conjugation,

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The solid supports with beads and immobilized molecules can be used in any application for which solid supports with immobilized molecules are used. For example, the methods and compositions described herein, can be used to isolate (purify) target nucleic acids from 5 biological samples (reactions). For example, the compositions and methods can be used to isolate particular nucleic acids, which are generated by cloning (Sambrook et al., Molecular Cloning : A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989), polymerase chain reaction (PCR) (C.R. Newton and A. Graham, PCR, BIOS Publishers, 10 1994), ligase chain reaction (LCR) (see, e.g., Wiedmann et al. (1994) PCR Methods Appl. 3:57-64; Barany (1991) Proc. Natl. Acad. Sci. U.S.A. 88:189-93), strand displacement amplification (SDA) (see, e.g., Walker et al. (1994) Nucl. Acids Res. 22:2670-77; European Patent Publication Number 0 684 315 entitled "Strand Displacement 15 Amplification Using Thermophilic Enzymes") and variations such as reverse transcriptase (RT)-PCR (Higuchi et al. (1993) Bio/Technology 11:1026-1030), allele-specific amplification (ASA), cycle sequencing and transcription based processes.

Further the methods and compositions can be used to isolate or 20 transfer particular nucleic acids during the performance of a particular reaction. For example, an amplification reaction, such as a PCR reaction, can be performed to 'master' mix without addition of the dideoxynucleotides (d/ddNTPs) or sequencing primers. Aliquots can be isolated via a conjugation means described herein and transferred, for 25 example to a sequencing plate, where d/ddNTPs and primers can then be

added between A, C, G, and T master mixes. Aliquots can then be transferred to a sequencing plate and sequencing primers added.

specificity, the pin tool (with or without voltage), can be modified to contain a partially or fully single stranded oligonucleotide (e.g., about 5-12 base pairs). Only complementary nucleic acid sequences (e.g. in solution) are then specifically conjugated to the pins.

5 In yet a further embodiment, a PCR primer can be conjugated to the tip of a pin tool. PCR can be performed with the solid phase (pin tool)-bound primer and a primer in solution, so that the PCR product becomes attached to the pin tool. The pin tool with the amplification product can then be removed from the reaction and analyzed (e.g., by
10 mass spectrometry).

Examples of different pin conformations are shown in Figure 9. For example, Figures 9a, 9b and 9c show a solid pin configuration. Figures 9d and 9e show pins with a channel or hole through the center, for example to accommodate an optic fiber for mass spectrometer
15 detection. The pin can have a flat tip or any of a number of configurations, including nanowell, concave, convex, truncated conic or truncated pyramidal (e.g. size 4-800 μ across x 100 μ depth). In a preferred embodiment, the individual pins, which can be any desired size, are preferably up to about 10 mm in length, and more preferably are
20 about 5 mm in length and about 1 mm in diameter. The pins and mounting plate can be made of polystyrene (e.g. one-piece injection molded). Polystyrene is an ideal material to be functionalized and can be molded with very high tolerances. The pins in a pin tool apparatus may be collapsible (e.g., controlled by a scissor-like mechanism), so that pins
25 may be brought into closer proximity, reducing the overall size.

Scans including, for example, spectrometric techniques such as UV/VIS, IR, fluorescence, chemiluminescence, or NMR spectroscopy, mass spectrometry, or other methods known in the art, or combinations

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Two mass spectrometer geometries for accommodating the pin tool apparatus (see Figure 9) are proposed in Figure 13. The first accommodates solid pins. In effect, the laser ablates a layer of material from the surface of the crystals, the resultant ions being accelerated and 5 focused through the ion optics. The second geometry accommodates fibre optic pins in which the samples are lasered from behind. In effect, the laser is focused onto the pin tool back plate and into a short optical fibre (about 100 μm in diameter and about 7 mm length to include thickness of the back plate). This geometry requires the volatilized 10 sample to go through the depth of the matrix/bead mix, slowing and cooling down the ions resulting in a type of delayed extraction which should actually increase the resolution of the analysis.

The probe through which the pins are fitted can also be of various geometries. For example, a large probe with multiple holes, one for each 15 pin, fitted over the pin tool. The entire assembly is translated in the X-Y axes in the mass spectrometer. Alternatively, as a fixed probe with a single hole, which is large enough to give an adequate electric field, but small enough to fit between the pins. The pin tool is then translated in all three axes with each pin being introduced through the hole for 20 sequential analyses. This format is more suitable for the higher density pin tool (i.e. based on a 384 well or higher density microplate format). The two probes described above, are suitable for the two mass spectrometer geometries described above.

Figure 14 schematically depicts the steps involved in mass 25 spectrometry sequencing by post biology capture as described above

parallel analysis of a single well

The methods provided herein are useful for providing spatially-addressable arrays of nucleic acids immobilized on beads, which are

disposes the sample arrays for analysis by mass spectrometry. A mass spectrometer is provided that generates a set of spectra signal that is indicative of the composition of the sample material under analysis.

In particular, the pin tool includes a housing having a plurality of sides and a bottom portion having formed therein a plurality of apertures, the walls and bottom portion of the housing defining an interior volume; one or more fluid transmitting vesicles, or pins, mounted within the apertures, having a nanovolume sized fluid holding chamber for holding nanovolumes of fluid, the fluid holding chamber being disposed in fluid communication with the interior volume of the housing, and a dispensing element that is in communication with the interior volume of the housing for selectively dispensing nanovolumes of fluid from the nanovolume sized fluid transmitting vesicles when the fluid is loaded with the fluid holding chambers of the vesicles. This allows the dispensing element to dispense nanovolumes of the fluid onto the surface of the substrate when the apparatus is disposed over and in registration with the substrate.

In one embodiment, the fluid transmitting vesicle has an open proximal end and a distal tip portion that extends beyond the housing bottom portion when mounted within the apertures. In this way the open proximal end can dispose the fluid holding chamber in fluid communication with the interior volume when mounted with the apertures. Optionally, the plurality of fluid transmitting vesicles are removably and replaceably mounted within the apertures of the housing, or alternatively can include a glue seal for fixedly mounting the vesicles.

In another embodiment, the fluid holding chamber includes a narrow bore dimensionally adapted for being filled with the fluid through capillary action, and can be sized to fill substantially completely with the

housing at varying pressure conditions. This allows the controller varying element to dispose the interior volume at a selected pressure condition sufficient to offset the capillary action to fill the fluid holding chamber of each vesicle to a predetermined height corresponding to a 5 predetermined fluid amount. Additionally, the controller can further include a fluid selection element for selectively discharging a selected nanovolume fluid amount from the chamber of each the vesicle. In one particular embodiment, the a pressure controller that operates under the controller of a computer program operating on a data processing system 10 to provide variable control over the pressure applied to the interior chamber of the housing is provided.

The fluid transmitting vesicle can have a proximal end that opens onto the interior volume of the housing, and the fluid holding chamber of the vesicles are sized to substantially completely fill with the fluid 15 through capillary action without forming a meniscus at the proximal open end. Optionally, the apparatus can have plural vesicles, wherein a first portion of the plural vesicles include fluid holding chambers of a first size and a second portion including fluid holding chambers of a second size, whereby plural fluid volumes can be dispensed.

20 In another embodiment, the tool can include a fluid selection element that has a pressure source coupled to the housing and in communication with the interior volume for disposing the interior volume at a selected pressure condition, and an adjustment element that couples to the pressure source for varying the pressure within the interior volume 25 of the housing to apply a positive pressure in the fluid chamber of each

theretrom. The selection element and adjustment element can be computer programs operating on a data processing system that directs the operation of a pressure controller connected to the interior chamber

Kits

Also provided is a kit for immobilizing nucleic acids on beads, which are further bound. In one embodiment, the kit includes an appropriate amount of: i) beads, and/or ii) the insoluble support, and iii) 5 conjugation means. The kits described herein can also optionally include appropriate buffers; containers for holding the reagents; and/or instructions for use.

The following examples are included for illustrative purposes only and are not intended to limit the scope of the invention.

10

EXAMPLE 1**Attachment of Resin Beads to a Silicon Surface**

A silicon surface (e.g. of a silicon wafer) is derivatized with amino groups by treatment with 3-aminopropyltriethoxysilane. Wang resin beads are treated with succinic anhydride to provide carboxyl-15 functionalized resin beads. The carboxyl-functionalized resin beads are then coupled to the amino-functionalized silicon surface with a coupling reagent (for example, dicyclohexylcarbodiimide (DCC)), in the presence of p-nitrophenol. The resin beads become covalently linked to the silicon surface, and the unreacted carboxyl groups of the resin are converted to 20 the p-nitrophenyl ester (an activated ester suitable for coupling with a nucleic acid).

Alternatively, the carboxyl groups of the Wang resin are transformed to the p-nitrophenyl active esters prior to reacting with the amino-functionalized silicon surface.

25 Thus, resin beads can be rapidly and conveniently attached to a

suitable for covalent attachment of nucleic acids.

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Alternatively C18 beads were first covalently attached to a silicon surface (e.g. a silicon wafer) or adsorbed to a flat surface by hydrophobic interaction.

The results showed that acetonitrile/water at levels of ca. > 30%
5 are enough to dissociate the hydrophobic interaction. Since the matrix used in MALDI contains 50% acetonitrile, the DNA can be released from the support and MALDied successfully (with the trityl group removed during the MALDI process).

EXAMPLE 4

10 Attaching Beads to Silicon Chips

Amino derivatization of silicon surface

Silicon wafers were washed with ethanol to remove surface debris and flamed over a bunsen burner until "red hot" to ensure oxidation of the surface. After cooling, the wafers were immersed in an anhydrous
15 solution of 3-aminopropyltriethoxysilane in toluene (25% v/v) for 3 hours. The wafers were then washed with toluene (three times) then anhydrous dimethylacetamide (three times).

Activation of Wang resin beads

Vacuum-dried Wang resin beads (5 g, 0.84 mmol/g loading, 4.2
20 mmol, diameter 100-200 mesh), obtained from Novabiochem, were suspended in pyridine (40 ml) with DMAP (0.1 eq, 0.42 mmol, 51 mg). To this was added succinic anhydride (5 eq, 21 mmol, 2.10 g) and the reaction was shaken for 12 hours at room temperature. After this time, the beads were washed with dimethylformamide (three times), then
25 pyridine (three times) and suspended in pyridine/dimethylformamide (1:1

Condensation was activated by adding dicyclohexylcarbodiimide (DCC) (2 eq, 8.4 mmol, 1.73 g) and the reaction mixture was shaken for 12

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and 0.5 μ l of supernatant was applied to MALDI-TOF MS. To maximize the recovery of the bound iminobiotin oligo, the beads from the above treatment were again incubated with 2 μ l of 3-HPA matrix and 0.5 μ l of the supernatant was applied to MALDI-TOF MS.

5 Matrix alone and free biotin treatment quantitatively released iminobiotin oligo off the streptavidin beads as shown in Figures 5 and 6. Almost no bound oligo was observed after the second treatment which confirmed the complete recovery.

Equivalents

10 Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, numerous equivalents to the specific procedures described herein. Such equivalents are considered to be within the scope of this invention and are covered by the following claims.

15 Since modifications will be apparent to those of skill in this art, it is intended that this invention be limited only by the scope of the appended claims.

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- (A) TELEPHONE: 619-238-0999
- (B) TELEFAX: 619-238-0062
- (C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TGCACCTGAC TC

12

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TGTGCGTCGT G/

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11. A process of making a bead conjugated to a solid support and further conjugated to a macromolecule or biological particle, comprising:

(a) conjugating a bead to a macromolecule or biological particle;
5 and
(b) conjugating a bead to a solid support, wherein steps (a) and (b) are performed sequentially in any order or simultaneously.

12. A process of claim 11, wherein the macromolecule is selected from the group consisting of nucleic acids, peptides, proteins,
10 amino acids and organic molecules.

13. A process of claim 11, wherein the bead is conjugated to a macromolecule and the macromolecule is a nucleic acid.

14. A process of any of claims 11-13, wherein the bead is functionalized for linkage of macromolecules.

15. A process of any of claims 11-13, wherein the bead is functionalized with carboxy functional groups.

16. A process of any of claims 11-13, wherein the bead is functionalized with amino functional groups.

17. A process of any of claims 11-13, wherein the bead is conjugated to the macromolecule or biological particle prior to conjugation of the bead to the solid support.

18. A process of claim 10, wherein the bead is conjugated to the macromolecule or biological particle after the bead is conjugated to the solid support.

25 19. A kit, comprising:

i) the insoluble support, and
ii) conjugation means for linking molecules to the beads and the beads to the support.

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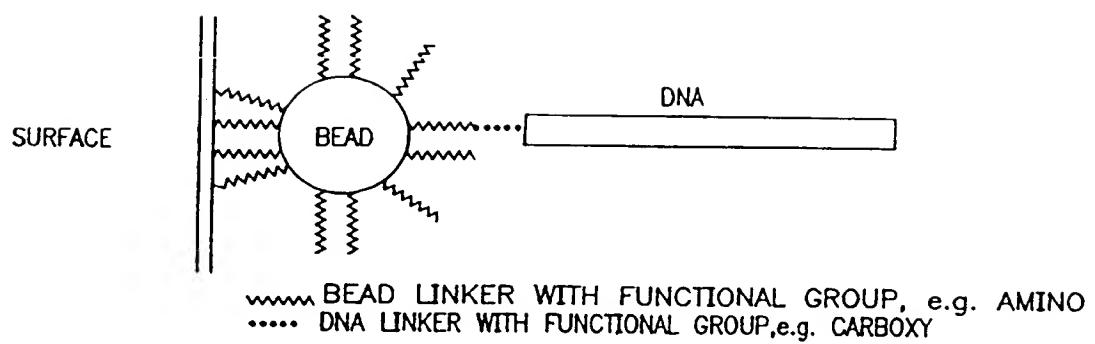


FIG. 1

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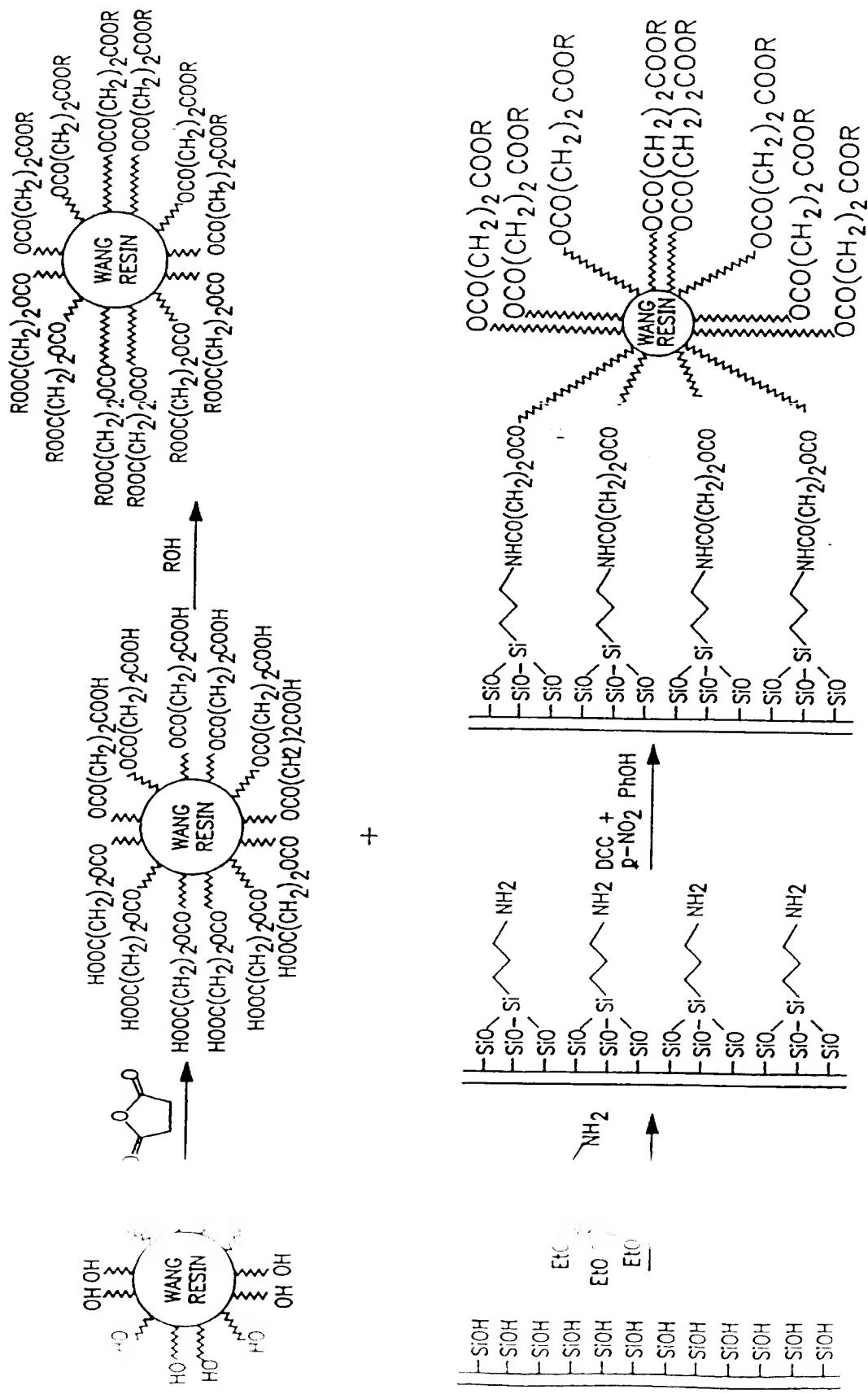
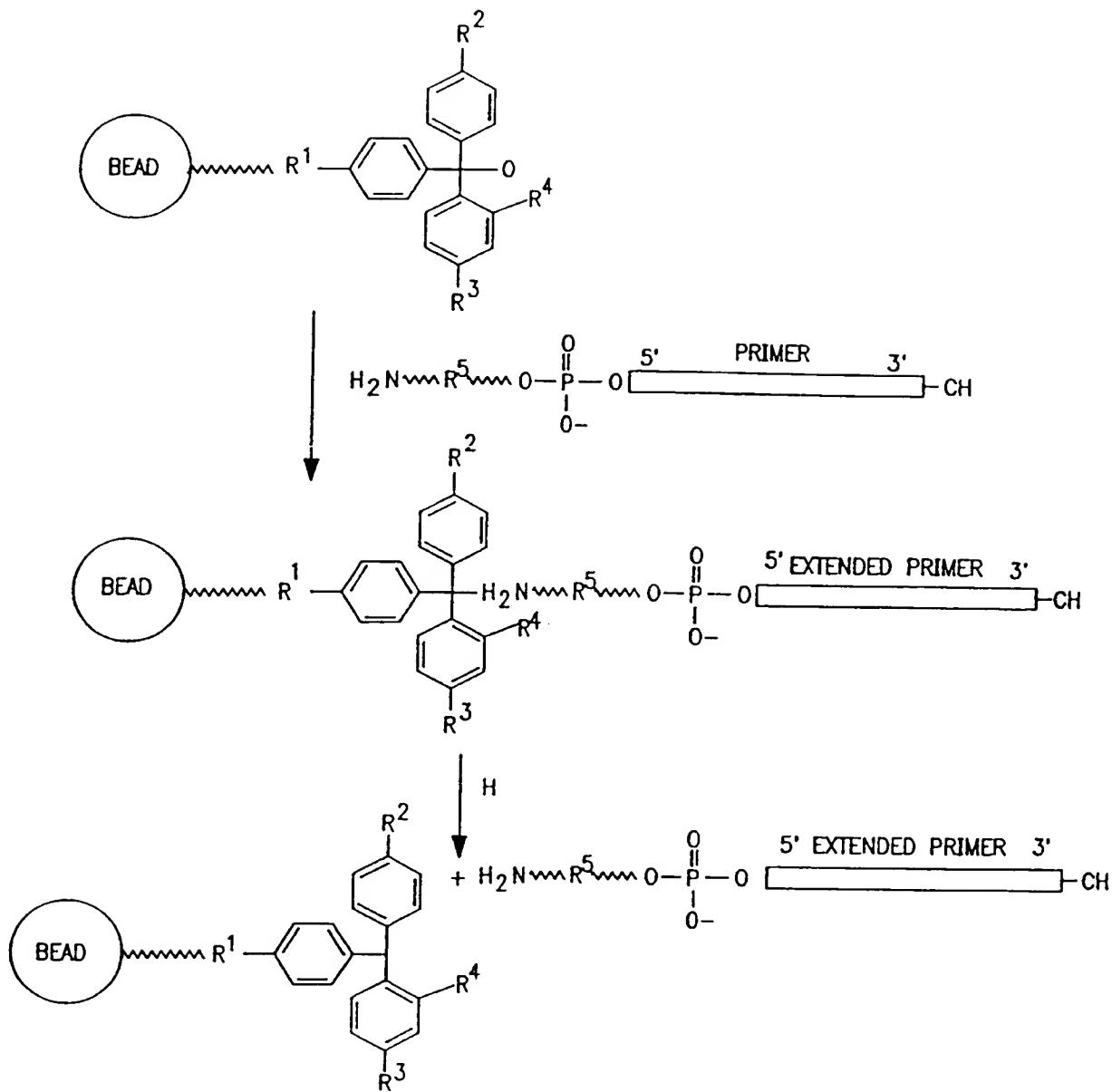


FIG. 2

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$R^1 = COO(CH_2)$; para or meta
 $R^2 = MeO; H$
 $R^3 = MeO; H$

FIG. 3

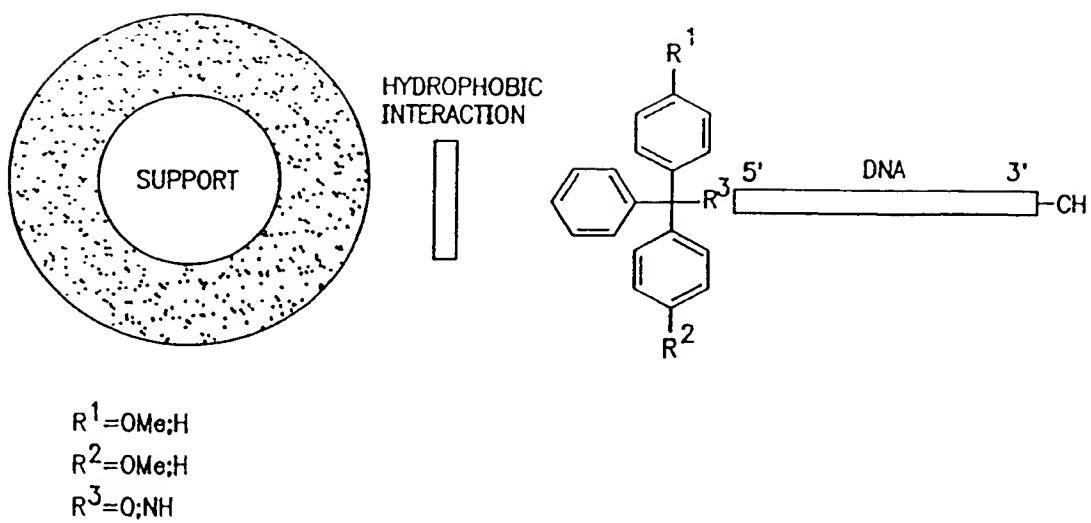


FIG. 4

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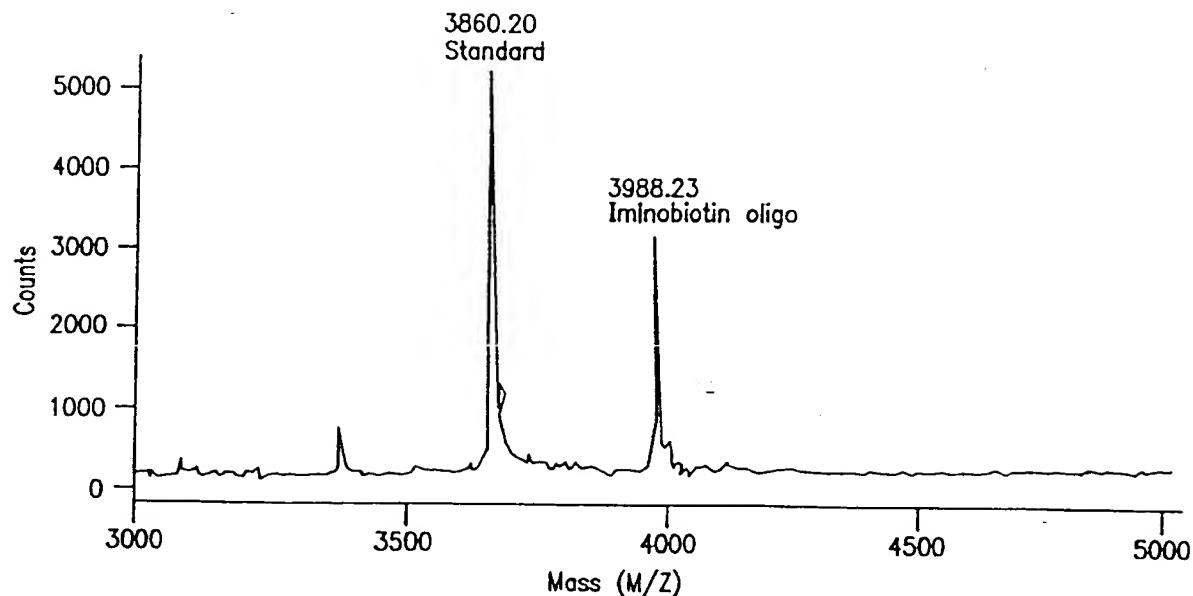


FIG. 5

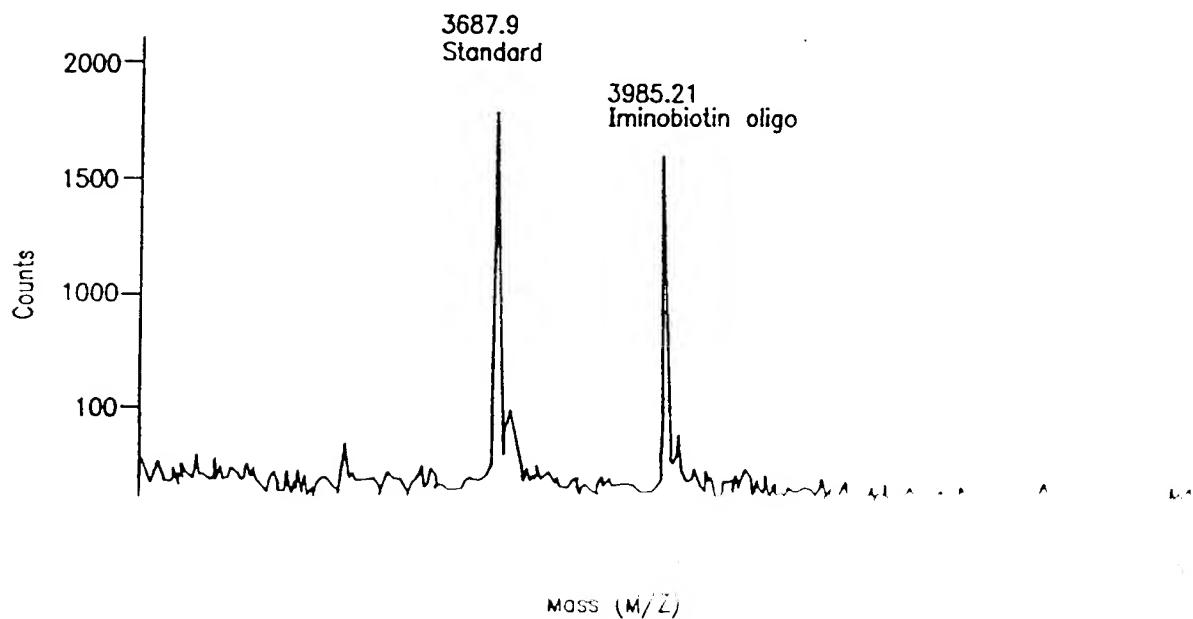


FIG. 6

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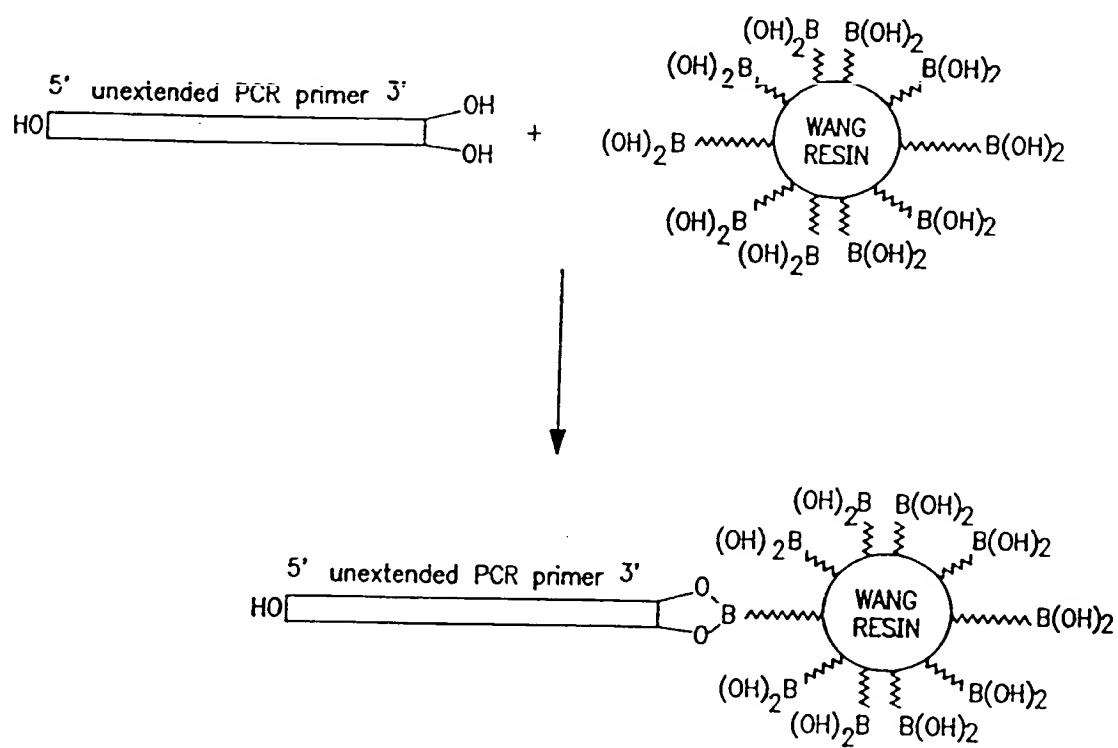


FIG. 7

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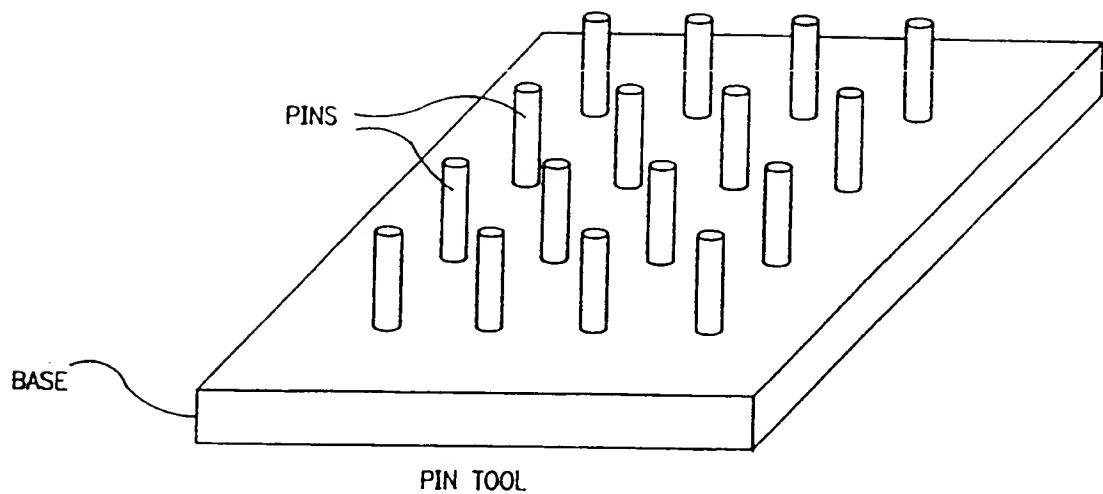


FIG. 8

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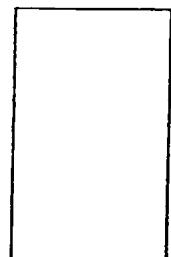


FIG. 9A

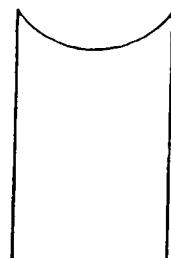


FIG. 9B

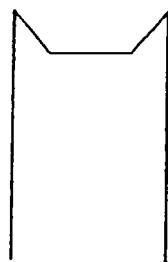


FIG. 9C

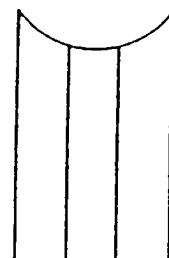


FIG. 9D

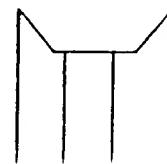


FIG. 9E

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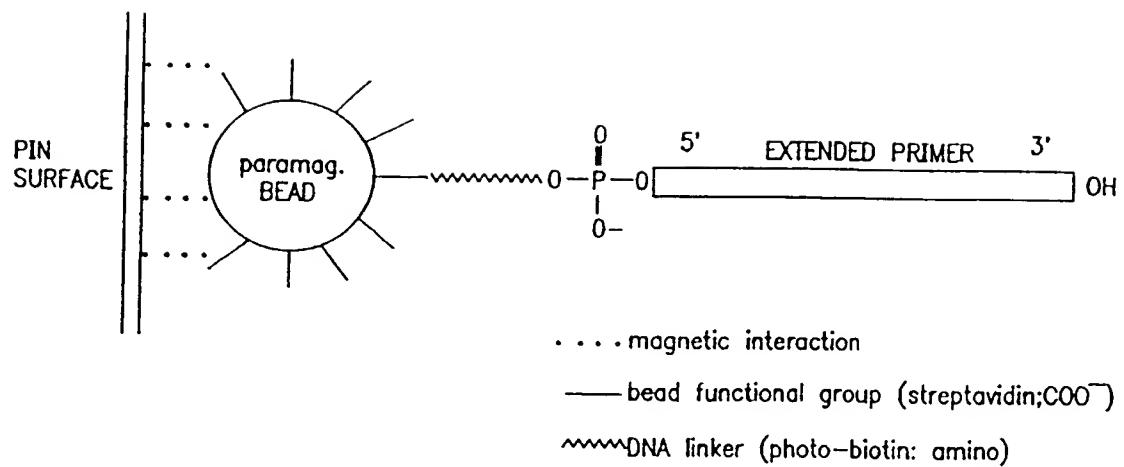
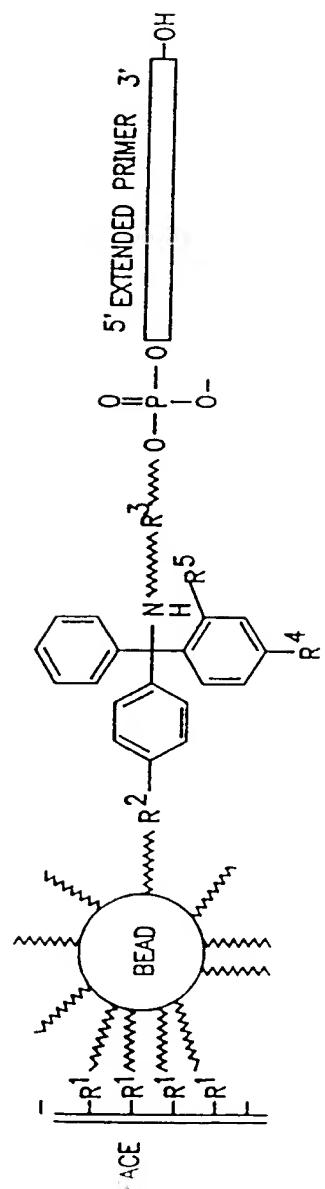


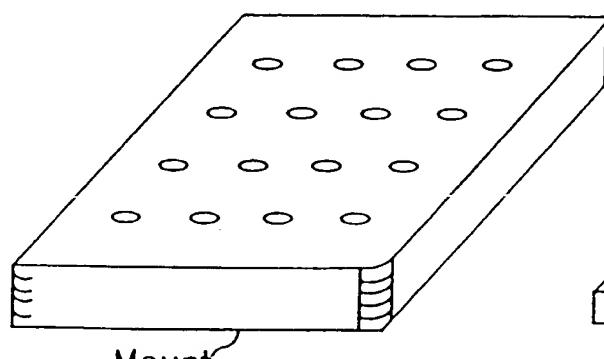
FIG. 11



$R^1 = (CH_2)_a NHCO(CH_2)_b : (CH_2)_c S - S(CH_2)_d$
 $R^2 = (CH_2)_e CONH(CH_2)_f : (CH_2)_g S - S(CH_2)_h$
 $R^3 = MeO; H$
 $R^4 = MeO; H$
 $R^5 = Cl; H$
 $R^6 = (CH_2)_m : (CH_2)_n CONH(CH_2)_p$

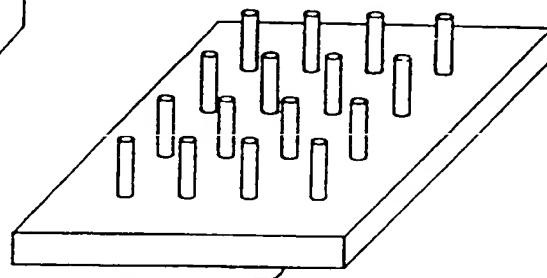
FIG. 10

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Mount

FIG. I2A



Pin tool

FIG. I2B

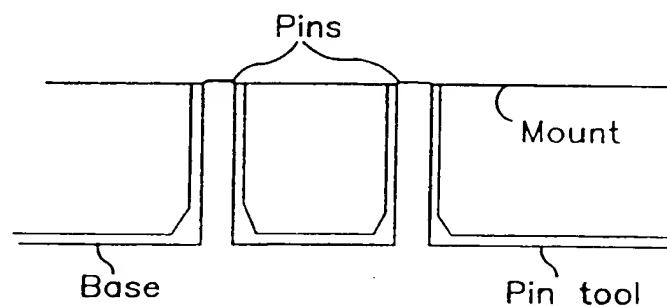


FIG. I2C

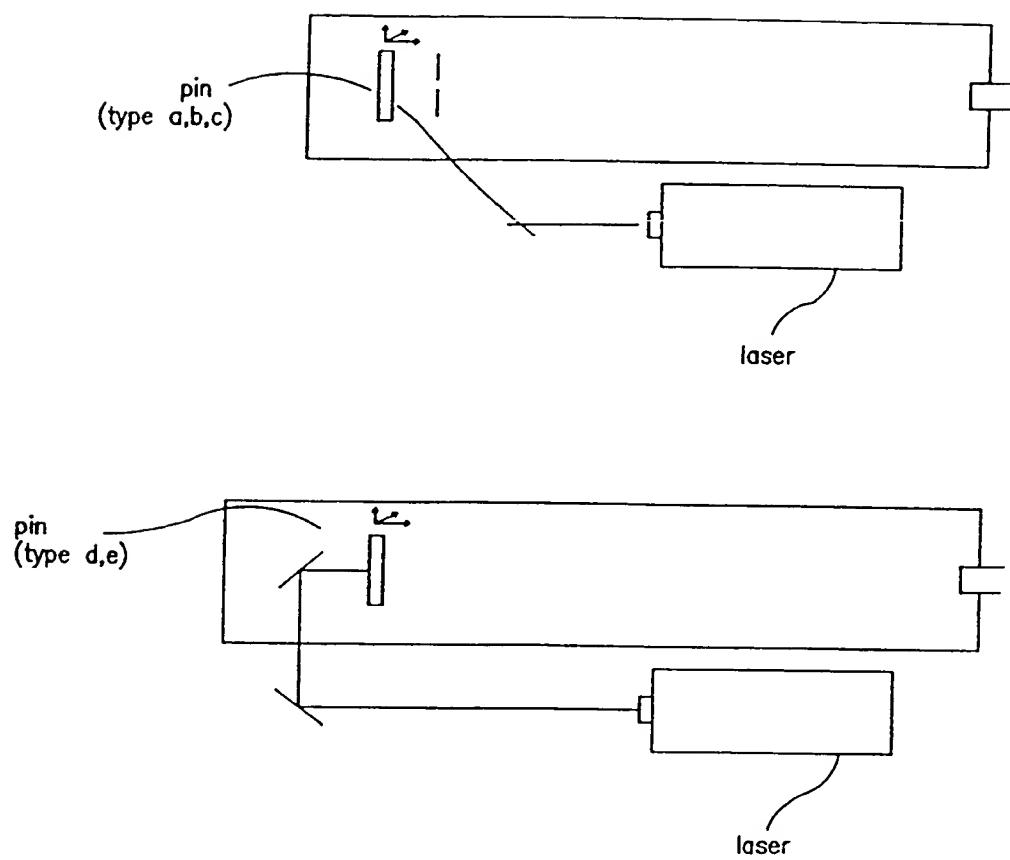


FIG. 13

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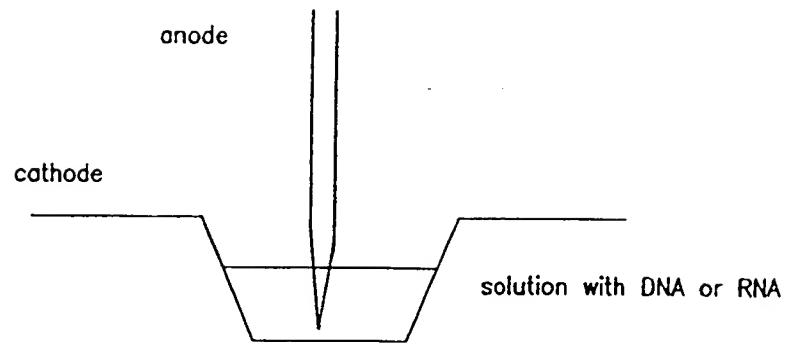


FIG. 14

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Experimental flow diagram:

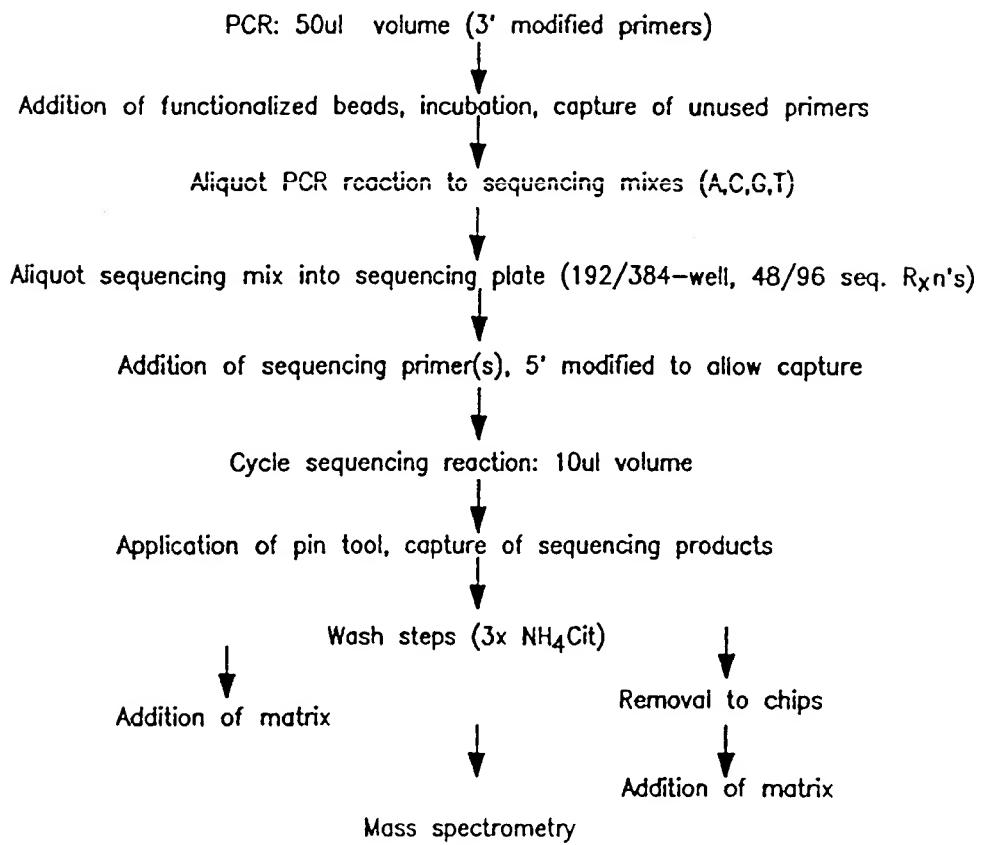


FIG. 15

INTERNATIONAL SEARCH REPORT

Int'l. Application No.
PCT/US 97/20194

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C07H21/00

According to International Patent Classification(IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C07H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category * | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|------------|--|-----------------------|
| X | EP 0 455 905 A (MICROPROBE CORP) 13 November 1991 see abstract see page 2, line 55 – page 3, line 17 see page 3, line 31 – line 51 see page 7, line 22 – line 39 see claims 1-41 — | 1-19 |
| X | WO 93 06925 A (MINNESOTA MINING & MFG) 15 April 1993 see the whole document — | 1-19 |
| X | ARSHADY R: "BEADED POLYMER SUPPORTS AND GELS I. MANUFACTURING TECHNIQUES" JOURNAL OF CHROMATOGRAPHY, vol. 586, no. 2, 22 November 1991, pages 181-197, XP000247968 see the whole document — | 1-10 |
| | | -/- |

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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Date of the actual completion of the international search

20 March 1998

Date of mailing of the international search report

27/03/1998

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Scott, J

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int'l Application No
PCT/US 97/20194

| Patent document cited in search report | Publication date | Patent family member(s) | Publication date |
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| WO 9306925 A | 15-04-93 | CA 2114732 A DE 69223798 D EP 0607327 A JP 7500363 T | 15-04-93 05-02-98 27-07-94 12-01-95 |
| US 4797355 A | 10-01-89 | NONE | |
| EP 0396116 A | 07-11-90 | CA 2015938 A DE 69029873 D DE 69029873 T ES 2099699 T JP 2304364 A US 5399501 A | 02-11-90 20-03-97 07-08-97 01-06-97 18-12-90 21-03-95 |